https://doi.org/10.46344/JBINO.2022.v11i01.15

PHYTOCHEMICAL ASSAY OF LEAF AND ROOT OF METHANOL EXTRACTS OF THAUMATOCOCCUS DANIELLI

EBULUE, M.M.

Department of Biotechnology, Federal University of Technology Owerri, Imo State, Nigeria

ABSTRACT

Phytochemicals are the active components in most plants that contribute to their protective activity. This study quantitatively determined the phytochemical composition of dried leaves and roots of *Thaumatococcus danielli*. Phytochemical analysis using GC-FID (BUCK M910) revealed that kaempferol was higher in leaf (18.75ug/g) than in root (9.55ug/g), oxalate was higher in leaf (1.2ug/g) than in root (0.84ug/g), anthocyanin (2.49ug/g) higher in leaf than in root (1.37ug/g), saponin (14.33ug/g) higher in leaf than in root (14.00), phenol was higher in leaf (6.66ug/g) than in root (3.96ug/g). While tannin was higher in root (10.39ug/g) than in leaf (8.93ug/g), Lunamarine was higher in root (2.08uu/g) than in leaves (1.98ug/g) and ribalinidine was higher in root (4.76ug/g) than in leaf (3.09ug/g). Qualitative analysis revealed significantly presence of flavonoid (++), saponin (++) and tannin (++) in both leaf and root extracts. Alkaloids, proteins and cardiac glycosides were found in trace amounts (+) while terpenoids, resins and steroids were not found (-) in both extracts and these account for the plant's medicinal value as antidote against venoms, stings and bites and sedative for treating insanity.

KEY WORDS: Thaumatococcus danielli (leaf, root), Phytochemicals





INTRODUCTION

The active components in most plants that contribute to their protective effects are the phytochemicals, vitamins and minerals (Okwu, D.E. and Ekeke, O. 2003). Plants are being used for numerous effects beneficial nutrition in therapeutics (protective and medicinal) (Assareh et al., mankind Beneficial effects result from the mix of secondary metabolites that are capable of generating physiological actions within the body. Thaumatococcus danielli is a multipurpose, monocotyledonous tropical plant found in the rainforest region of West Africa particularly Nigeria and Ghana (Yeboah et al., 2003; Ojekale et al., 2007).

In Nigeria, the leaves of this plant are used among different ethnic groups for packaging and preservation of cooked foods such as meat, rice, beans, maize meal and beans cake etc. A sweet protein, Thaumatin, which is the source of natural sweetener, is found in the fruit of this plant, Thaumatococcus danielli (Arowosege and Labode, 2006). The fruit of this plant is used as laxative and due to its non-caloric value it has also been shown to be very ideal in the management of diabetes mellitus while the use of seed in emesis for pulmonary disorder, the leaf sap as antidote against stings venoms, and (Emudainohwo et al., 2015) and the root and leaf sap as sedative for treating insanity (Shalom et al., 2014) informs this research to evaluate the bioactive compounds (phytochemicals) it contains.

Objectives of study

It is to determine the qualitative and quantitative phytochemical compositions

of root and leaf of methanol extracts of Thaumatococcus danielli.

Materials and methods Collection of plant materials

Fresh leaves and roots of Thaumatococcus danielli were collected from the bush, identified in Plant Science and Biotechnology Department of Federal University Technology Owerri, Imo State, Nigeria. The roots and leaves (samples) were shade-dried at room temperature to a constant weight, ground to powder, packed into polythene bags and stored for subsequent uses.

Sample extraction

About 200g of each powdered sample (leaves and roots) was macerated in 1L of methanol at room temperature for 72hrs, filtered with Whatmann No. 1 filter paper and the filtrate was transferred into a rotary evaporator at 40 – 42°C. Each residue obtained was further dried in a water bath at 37 – 40°C. The crude extracts were stored in a refrigerator at 4°C.

Qualitative and quantitative phytochemical analysis

Qualitative analysis was carried out using the method of Trease and Evans and Harbone (1998) to identify the different phytochemicals, while phytochemical quantification was performed on a BUCK M910 GC equipped with flame ionization detector.

2022 January Edition | www.jbino.com | Innovative Association

RESULT

Table 1: Result of qualitative phytochemical screening

| Parameters | Leaf | Root |
|----------------------------|------|------|
| Alkaloids | + | + |
| Flavonoids | ++ | ++ |
| Steroids | - | - |
| Tannins | ++ | ++ |
| Resins | - | - |
| Proteins | + | + |
| Saponins | ++ | ++ |
| Cardiac glycosides | + | + |
| Terpenoids | - | - |
| (+) trace amount | | |
| (++) significantly present | | |
| (-) absence | | |

Table 2: Result of quantitative phytochemical analysis using GC-FID

| Phytochemicals | Leaf ug/g | Root ug/g |
|----------------|-----------|-----------|
| | | |
| Phytate | 0.23 | 0.29 |
| Epidcatechin | 1.80 | 1.87 |
| Oxalate | 1.20 | 0.84 |
| Anthocyanin | 2.49 | 1.37 |
| Tannin | 8.93 | 10.39 |
| Phenol | 6.66 | 3.96 |
| Lunamarine | 1.98 | 2.08 |
| Ribalinidine | 3.09 | 4.76 |
| Catechin | 5.51 | 5.67 |
| Rutin | 11.25 | 11.35 |
| Kaempferol | 18.57 | 9.55 |
| Saponin | 14.33 | 14.00 |
| Spartein | 0.0002 | 0.0002 |
| | | |

DISCUSSION

Phytochemicals such as flavonoids and phenols in line with other antioxidants, functions within the body by mopping up free radicals and metallic ions. Flavonoids and phenols are the largest group of phytochemicals that account antioxidants activity in plants or plant material (Gupta et al., 2012). These antioxidants are capable of slowing or preventing the oxidation of other molecules. The uncontrolled production of free radicals is associated with the onset of numerous diseases like cancer. rheumatoid arthritis, as well as in the degenerative process associated with Parkinson's including aaina, Alzheimer's diseases (Ali et al., 2008; Di Matteo and Esposito 2003).

Phytochemical analysis revealed high level of flavonoids in both leaves and roots of which kaempferol concentration was much higher in leaves than in roots. Tannin was higher in roots than in leaves, while phenol was higher in leaves than in roots. The level of phytate and oxalate in both samples were very low, but saponin was significantly present in both leaves and roots. Lunamarine and Ribalinidine were higher in roots than in leaves and this account for its medicinal value.

CONCLUSION

The phytochemical analysis of this plant showed significant concentrations flavonoids, a powerful antioxidant in conjuction with other phytochemicals which have physiological role in mopping up free radicals. Thus the plant could also in the prevention be used and management of degenerative diseases such as diabetes mellitus and

cardiovascular disorders.

REFERENCES

Ali, S.S., N. Kasoju, A. Luthra, A. Singh, H. Sharanabasava (2008). Indian Medicinal Herbs as Sources of Antioxidants. Food Res. Intl., 41(1): 1-15.

Arowosoge,O.G.E and Labode, (2006). Economic analysis of Thaumatococcus daniellii (Benn.) benth. (Miraculous berry) in Ekiti State, Nigeria. Journal of Food Agriculture & Environment, 4(1): 264-269.

Assareh MH, Sedaghati M, Kiarostami K, Zare AG. (2010). Seasonal changes of essential oil composition of Eucalyptus maculata Hook. Iranian Journal of Medicinal and Aromatic Plants, **25**: 580-588.

Di Matteo V. and Esposito E. (2003) Biochemical and therapeutic effects of antioxidants in the treatment of Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis. Curr Drug Targets CNS Neurol Disord, 2(2): 95-107.

Emudainohwo, J.O.T., Erhirhie, E.O., Moke, E.G. and Edje, K.E. (2015). A Comprehensive Review on Fthno-Medicine, Phytochemistry and Ethnopharmacology of Chrysophyllum albidum Journal of Advances in Medical and Pharmaceutical Sciences, 3(4): 147-154.

Gupta, S., Kumar, M.N.S., Duraiswamy, B., Chhajed, M. and Chhajed, A. (2012). In-

2022 January Edition | www.jbino.com | Innovative Association



vitroAntioxidant and Free Radical Scavenging Activities of Ocimum Sanctum. World Journal of Pharmaceutical Research, 1: 78-94.

Harborne, J.B. Phytochemical methods. A guide to modern technique of plant analysis (3rd edn). Chapman and Hall, London, 1998; 88-185.

Ojekale, A.B, Makinde, S.C.O. and Osileye, O. (2007). Phytochemistry and antimicrobial evaluation of Thaumatococcus danielli, Benn. (Benth.) leaves. Nigerian Food Journal, DOI: 10.4314/nifoj.v25i2.50858.

Okwu, D.E. and Ekeke, O. (2003). Phytochemical screening and mineral composition of chewing sticks in south Eastern Nigeria. *Global journal of Pure and Applied Science*, **9**: 235-238.

Shalom, N. C., Adebayo, Y. O, Samuel, T. P., Bolaji, J. D. and Tamunotonyesia, E. (2014). Analysis of leaf, fruit and seed of Thaumatococcusdanielli (Benth). Pakist an Journal of Biological Sciences 17 (6) 849-854.

Trease, G.E, Evans MC. Textbook on Pharmacognosy (13th Edn). Bailiere Tandal and Caussel, London, 1989; 144-148.

Yeboah, S. O., Hilger, T.H. and Kroschel, J. (2003). *Thaumatococcus danielli* (Benn.) Benth, A Natural Sweetener from the Rain Forest Zone in West Africa with Potential for Income Generation in Small Scale Farming. Institute of Plant Production and Agroecology of the Tropics and

Subtropics, Hohenheim University, Stuttgart (Germany), Email: t-hilger@uni-hohenheim.de, Fax: +49[**0**]711-4592304.

