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EFFECT OF THIOBARBIUTRIC ACID ACTIVITY IN PLASMA OF DIABETIC PATIENTS WITH NEPHROPATHY – A Review

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ABSTRACT

The worldwide incidence of diabetes has increased dramatically along with widespread lifestyle and dietary changes. Diets high in fat are strongly associated with the development of obesity and can induce insulin resistance in humans and animals. It is clear that obesity constitutes a risk factor for contributing to the development of type 2 diabetes. In the present study, we investigated the therapeutic potential action of Thiobarbeutric acid on diabetes associated complications.

Key words: Thio Barbeutic acid, Diabetes retinopathy.

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INTRODUCTION

Diabetes is a serious disease characterized insufficient insulin secretion, insulin by activity deficiency and hyperglycaemia, estimated prevalence is pretty and its high (5.4% in 2025) in society. At the same time, it causes to serious problems involving blindness, pancreatic cancer and impairment. coanitive Moreover, the molecular level effects of diabetes also exist and they involve increase in oxidative deficit levels and in antiagent oxidative systems. particular In hyperglycaemia in diabetes causes increase of oxidative radicals and then the radicals lead to diabetic complications. Especially these oxidative radicals lead to structural defects of membranes and proteins. In the present study we are concentrating on effect of thiobarbeutric acid in plasma of diabetic patients with nephropathy

Discussion

We have earlier reported the hypoglycemic and hypo cholesterolemic activities of *K. pinnata* preparation consumption in streptozotocin-induced diabetes. This report provides data on the

role of K. pinnata preparation consumption on lipid peroxidation, antioxidant enzymes and RBC membrane ATPase activities in streptozotocin-induced diabetic rats. Diabetic conditions have been shown to result in impaired antioxidant defenses. compromised mitochondrial function, and increased sorbital and advanced alvcation end products from glucose. Previous studies have shown that oxidative stress aenerated plays hyperglycemia a particularly important role in the initiation of vascular diabetic complications, including retinopathy, cardiomyopathy, and nephropathy. SOD and CAT are antioxidant enzymes that convert toxic free radicals to water or other harmless compounds. Lipid peroxidation is intricately linked to antioxidant enzymes such as SOD and CAT. As a defense against reactive free radicals. the body produces antioxidant enzymes which help to prevent oxidative stress damage to tissues. The observed decreases in SOD and CAT activities and GSH levels in the untreated diabetic group is consistent with reports in the literature suggesting the disposition of



diabetic

diabetic tissues to damage bv free radicals Although, the TBARS levels were not significantly altered among the groups in this short-term study, the observed early increased SOD and CAT activities in treated diabetic rats with K. pinnata preparation may be indicative that the preparation might be effective in curtailing lipid peroxidation associated with disease. This would spare tissue damage and prevent the development of

complications.

Constant exposures to free radicals and high oxidative stress in diabetes have also been associated with erythrocyte structural damage. Lipid peroxidation alters cellular structure of membrane-bound enzymes by changing phospholipids and fatty acid composition. Previous studies have reported reduced erythrocyte **ATPase** activity, specifically Na +/K + ATPase in insulin-deficient conditions. In this study, we noted a nonsignificant decrease in Na +/K + and Mg²⁺ ATPase activities and nonsignificant increase in Ca²⁺ ATPase activity in the diabetic control group. However, in the diabetic group treated with aqueous K. pinnata preparation, there was a marginal increase in Na +/K + activity

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and a significant (P < 0.05) increase in Mg²⁺ ATPase activity compared to the diabetic control group. Mg²⁺ ATPase activity in the erythrocyte membrane has been reported to reduce cellular calcium content, improve erythrocyte flexibility and reduce vascular complications. In fact, in vivo studies vitro and in have demonstrated that insulin may modulate the shift of magnesium from extracellular to intracellular spaces. Intracellular Mg²⁺ is a critical cofactor for several enzymes in carbohydrate metabolism because of its role as part of the activated Mg²⁺ ATP The complex. activated complex responsible for the phosphorylation of all the rate limiting enzymes in the glycolytic pathway. Mg²⁺deficiency may also result in disorders of tyrosine-kinase activity on insulin receptor, leading to the development of post receptorial insulin resistance and decreased cellular glucose utilization. Hence, the observed significant increase in RBC membrane Mg²⁺ ATPase activity may suggest that the consumption of aqueous K. pinnata preparation could increase intracellular magnesium and subsequently improve rates of glycolytic activity. Adamson et al. also reported that the determination of ATPases, particularly Mg²⁺ ATPase and Na⁺/K⁺ ATPase, may



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Glutathione-S-transferases:

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provide an indirect measurement of insulin binding or level. We hypothesize that the consumption of K. pinnata preparation may act to increase peripheral insulin levels via extra pancreatic insulin elevation. It is also possible that the preparation's active ingredient may modulate the insulin-insulin receptor complex with a resultant prolonged half-life of ligandreceptor complex, which could lead to sustained signaling as was reported for the AspB10 insulin analog. More investigations are needed to further isolate and identify the active component(s) of the preparation, as well as to investigate, the effects of these isolates on glucoseregulating hormones.

REFERENCES

Ellman GL. Tissue sulfhydryl groups. Arch Biochem Biophys., 82, 1959, 70-77.

Kakkar P, Das B, Viswanathan PN. A modified spectrophotometric assay of superoxide dismutase. Indian J Biochem Biophy., 21,1978, 130–132.

Sinha AK. Colorimetric assay of catalase. Anal Biochem. 47, 1972, 389-394.

AB. Selenium: biochemical role as a component of glutathione peroxidase.

Levy Y, Zaltzberg H, Ben-Amotz A, Kanter Y, Aviram M. â-Carotene affects antioxidant status in non-insulin dependent diabetes mellitus. Pathophysiology 6, 1999, 157 – 161.

formation. J Biol Chem., 249, 1974, 7130-

Aydin A, Orhan H, Sayal A, Ozata M, Sahin G, Isimer A. Oxidative stress and nitric oxide related parameters in type II diabetes mellitus: effect of glycemic control. Clin Biochem., 34, 2001, 65–70.

Cho SY, Park JY, Park EM, Choi MS, Lee MY, Jeon SM, Jang MK, Kim MJ, Gupta RK, Kesari AN, Watal G, Murthy PS, Chandra R, Maithal K, Tandon V. Hypoglycemic and antidiabetic effect of aqueous extract of leaves of Annona squamosa. Curr Scien., 88, 2005,1244–1254.

Jeong HG, You HJ, Park SJ, Moon AR, Chung YC, Kana SK, Chun HK. Hepatoprotective effects of 18âglycyrrhetinic acid carbon on tetrachloride-induced liver injury: inhibition of cytochrome p450 2E1 expression.

Pharmacol Res., 46, 2002, 221-227.

Aebi H. Catalase in vitro. Methods Enzymol. 105, 1984, 121–126.

Jain SK, Levine SN. Elevated lipid peroxidation and vitamin E-quinone levels in heart ventricles of streptozotocin treated diabetic rats. Free Radic Biol Med., 18, 1995, 337–341.

Ingold KU, Webb AC, Witter D, Burton GW, Metcalf TA, Muller DP. Vitamin E remains the major lipid soluble, chain breaking antioxidant in human plasma even in individuals suffering from severe vitamin E deficiency. Arch Biochem Biophys., 259, 1987.224–225

Packer L, Tritschler HJ, Wessel K. Neuroprotection by metabolic antioxidant alpha lipoic acid. Free Radic Biol Med., 22,1997, 359–378.

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Freisleben HJ, Packer L. Free radical scavenging activities, interactions and recycling of antioxidants. Bio Soc Trans., 21, 1993, 325–333.

Garg M, Bansal DD. Protective antioxidant effect of vitamin C and vitamin E in STZ-induced diabetic rats. Indian J Exp Biol., 28, 2000,101–104.

Sajithlal GB, Chitra P, Chandrakasan G. Effect of curcumin on the advanced glycation and cross-linking of collagen in diabetic rats, Biochem Pharmacol., 56, 1998, 1607–1614.

