https://doi.org/10.46344/JBINO.2021.v10i06.27

TO STUDY PHYTOCHEMICAL SCREENING AND HPLC ANALYSIS PLANT EXTRACT OF LANTANA CAMARA

Dr. Amit C Lingayat

(Professor , Department of Dravyaguna, YCAMC, Aurangabad)

ABSTRACT

Since ancient time the herbal medicines are effective in the treatment of various ailments. Therefore, these plant drugs deserve detailed study in the light of modern science, and their taxonomical relatives can lead to the development of invaluable plant drugs for many dreadful diseases. In the present article we are discussing regarding the phytochemical and HPLC analysis of plant extract *Lantana camara*.

Key Words: Phytochemical Screening, lantana Camara.

INTRODUCTION

The essential values of some plants have long been published, however, a large number of them remain unexplored as yet. The most important of these bioactive constituents of plants are alkaloids, tannins, flavonoids, and phenolic compounds (Hill, 1952, Ali et al., 2001). Lantanoside, linaroside and camarinic acid have been isolated and are being investigated as potential nematocides (Day et al. 2003). Lantana oil is sometimes used for the treatment of skin itches, as an antiseptic for wounds, and externally for leprosy and scabies (Ghisalberti 2000). Plant extracts are used in folk medicine for the treatment of cancers, chicken pox, measles, asthma, ulcers, swellings, eczema, tumors, high blood pressure, bilious fevers, catarrhal infections, tetanus, rheumatism, malaria and atoxy of abdominal viscera (Ghisalberti 2000, Day et al. 2003). Lantana twigs and stems serve as useful fuel for cooking and heating in many developing countries (Sharma et al. 1988). Polyphenols especially TF exert cancer chemo preventive activity of inducing apoptic signals. (Lu et al., 1997, Yang et al., 2000, Javed et al., 1998). The anti inflammatory activity of the Bioactive Compound Oleanonic acid were tested against through the carrageen an induced Rat paw Oedema model (Ghosh et al., 2010)

In the present investigation, we report our findings on the total extractions of chemical components of the plant by HPLC method.

Materials and Methods

materials are collected Plant from university botanical aarden and the identified plant was confirmed by plant The preliminary taxonomist. phytochemical screening tests were carried out on the aqueous leaf crude extract of lantana Camara using standard procedures to identify the constituents.

Extraction of the Compound

For crude extraction fresh plant material were washed with tap water ,air dried and homogenized to fine powder and stored in air tight bottles . 10gms air dried powder mixed with 100 ml water at $37 \circ C$ for 48 Hrs .filtered in muslin cloth centrifuged at 500 g for 10 min. The supernatant was stored at 4 $^{\circ}C$

The compound can be extracted from above supernatant by passing through the column and is first fitted with Cotton and then silica gel, activated charcoal and again silica gel in ratio 1:2:1 and the crystals of plant extract are collected.

HPLC Analysis

The lantana Camara leaf extracted and purified compound was tested for the compound conformity and purity. The compounds extracted from leaf, stem, root and flower were set up for HPLC (High performance liquid chromatography) to test the purity of the compounds. In the HPLC mobile as well as stationary phases are used to test the purity of the compound. Usually a single gradient or a binary gradient are used as mobile phase. From the already available data it is known that the mobile phase used are shown in the figures itself. The detector used here is D₂ lamp as the measurement of the sample is <400 nm. The run time was set to 1 ml/1min. An injection volume of 20 ul is injected in to the stationary phase column. The gradient program was set up and the peak analysis was estimated by observing the graph and comparing the obtained chromatogram with that of the already available data (Plumb et al., 2004).

radicals by donating hydrogen atom or an electron to the free radical .It has been well known that plant materials have been well known that plant materials have shown to neutralize free radicals in various et invitro model systems (Zohang al.,1996).Earlier the poly phenolic compounds have protective effects on mutagenesis and carcinogenesis in human when ingested 1g daily from a diet rich in vegetables (Tanaka fruits and et al.,1989).The phenolic and flavonoid contents results of phytochemical analysis were carried out by HPLC method.

RESULT AND DISCUSSION

Phenolic compounds are reported to the active quenching oxygen –derived free

Qualitative Test	lantana
Camara	
Terpenes	Higher
Fixed Oils	Medium
Flavones	Higher
Alkaloids	low
Glycoside	Nil
Sterols	Nil
Phenols&Tannins	Nil

Table 1: Phyto Chemical Screening of lantana Camara

Bioactive potential of Flavonoids has been reported.(Illic et al.,2004 ,Cushner and Lamb 2005).The present results comparable with comparable with that of Samy and Ignace Muthu (1999).Sanchez et al (2005).

who reported the antimicrobial activity..In light of the fact that Micro organism are becoming resistant against the drugs in use , present investigation is of great importance in pharmaceutical industries for preparing plant based antimicrobial drugs.

REFERENCES

Ali, M. Rawinder E, Ramachandran R. (2001): A new flavonoid from the aerial parts of *Tridax* procumbens. Fitoterapia Mar 72; 3, 313-315.

Cushner TP, Lamb AJ (2005) .Antimicrobial activity of Flavonoids .Int. J.Antimicrobes Agents 26(5); 343-356

Day, M.D., Wiley, C.J., Playford, J. and Zalucki, P., (2003). Lantana: Current Management, Status and Future Prospects. Australian Centre for International Agricultural Research: Canberra.

Ghosh, S., Das Sarma, M., Patra, A. and Hazra, B. (2010), Anti-inflammatory and anticancer compounds isolated from Ventilago madraspatana Gaertn., Rubia cordifolia Linn. and Lantana camara Linn.Journal of Pharmacy and Pharmacology, 62: 1158–1166.

Ghisalberti EL., (2000). Lantana camara Linn. (Review). Fitoterapia. 71:467–485.

Hill AF (1952). Economic Botany. A textbook of useful plants and plant products. 2 nd edn. McGraw-Hill Book Company Inc, New York.

Ilic SB, Konstantinovic SS, Todorovic ZB. (2004). Antimicrobial activity of bioactive component from flower of Linum capitatum kit. Facta Universitatis 3(1): 73-78. Jane A, Plumb (2004): Cell sensitivity assays: The MTT assay ,Methods in Molecular Medicine, Volume 88, IV, 165-169, DOI: 10.1385/1-59259-406-9:165.

Javed S, Mehrotra NK and Shulka Y. (1998): Chempopreventive effects of black tea polyphenols in Mouse skin models of carcinogenesis. *Biomed. Environ. Sci*, 11:307-313

Lu YP, Lou YR, Xie JG, Yen P, Huang MT and Conney AH (1997): Inhibitary effectes of Black Tea on growth of eastablished skin tumour size, apoptosis, mitosis and bromo deoxo uridine incorporation into DNA. Carcinogenisis, 18:2163-2169.

RajuT.S,DavidSonE.A.,(1994).Carbhohydr.Res.258,243

Samy PR, Ignacimuthus S, Raja DP. (1999).Preliminary screening of ethnomedicinal plants from India. J.Ethanopharmacol 66(2): 235-240.

Sanchez SRP, Kantun SP, Tapia TWS, Pat FM, Polanco SP, Rivera RC. (2005). Screening of native plants from Yucatanfor anti- Giardia lamblia activity. *Pharm Biol* 43(7): 594-596

Sharma OP, Makkar HPS, Dawra RK., (1988); A review of the noxious plant Lantana camara. Toxicon. 26:975–987

Tanaka, Y., Bush, K. K., Klauck, T. M. & Higgins, P. J. (1989) Enhancement of butrayte -induced differentiation of HT-29 human colon carcinoma cells by way. Endrocrinology 136: 4157–4160.1,25dihydroxyvitamin D3. *Biochem. Pharmacol.* 38: 3859–3865. Yang GY, Liao J, Chung J, Yurkow EJ, Ho CT and Yang CS (2000): Effect of black and green tea polyphenols on c-jun phosphorylation and H₂O₂ production in transformed and non-transformed human bronchial cell lines: possible mechanisms of cell growth inhibition and apoptosis induction .Carcinogenesis ,21:2035-2039

Zhang L, Reith MEA (1996): Regulation of the functional activity of the human dopamine transporter by the arachidonic acid pathway. Eur J Pharmacol: 315:345– 354.